

Effects of Elevated O₃ and/or Elevated CO₂ on Lipid Peroxidation and Antioxidant Systems in *Ginkgo biloba* Leaves

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Received: 20 February 2008 / Accepted: 25 March 2009 / Published online: 7 April 2009
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Abstract Four-year-old seedlings of *Ginkgo biloba* were exposed to elevated O₃, elevated CO₂ and elevated O₃ plus elevated CO₂ in open-top chambers (OTCs) to study the responses of antioxidant system in *Ginkgo biloba* leaves. No significant changes in reactive oxygen production and scavenging systems were detected in seedlings exposed to high CO₂. Significant increase in H₂O₂ and MDA content were induced by elevated O₃. The ascorbate content and antioxidative enzymes activity were increased significantly by exposure to high O₃ as well. But the promoted ability in scavenging did not prevent the increase in H₂O₂ content and cell membrane lipid peroxidation. The increase was mitigated by high CO₂ in the combined exposure, but the effect was hardly significant.

Keywords Elevated O₃ and CO₂ · Antioxidant system

Troposphere concentrations of O₃ as well as CO₂ have been increasing rapidly as a result of human activities, and these two major greenhouse gases are likely to increase further, especially in urban areas of China (Wang et al.

2007). Tropospheric O₃ is a dangerous toxicant because it induces production of other reactive oxygen species (ROS) after diffusing into apoplastic solution. ROS were highly phytotoxic and would lead to oxidative stress in plant. Besides stomatal regulations, the resistance of plants to O₃ at cellular level is often correlated with the ability of cells to scavenge ROS by superoxide dismutase, catalase and other antioxidant enzymes in the ascorbate-glutathione cycle (Chernikova et al. 2000). Higher CO₂ may reduce the rate of ROS formation in photosynthesis apparatus for increased ratio of CO₂ to O₂. However, CO₂-driven down-regulation of detoxification system may make plants more susceptible to oxidative stress under higher CO₂ condition (Karnosky 2003). Both CO₂-induced stimulation (Schwanz et al. 1996) and down-regulation (Polle et al. 1993; Kull et al. 1996) of antioxidant systems have been reported. Results from recent years' studies on plant antioxidant systems in elevated O₃ and elevated CO₂ environment are still highly inconsistent though researches on the interactive effects of elevated O₃ and CO₂ on plant antioxidant systems have been started since the early 1980s (Polle et al. 1993; Tausz et al. 2004).

In the present work on *Ginkgo biloba*, which is one of the commonly used street trees for urban greening in Shenyang and other cities, hydrogen peroxide production, malondialdehyde (MDA) accumulation, ascorbic acid content and the specific activity of antioxidant enzymes were determined in leaves to investigate whether elevated CO₂ would ameliorate the oxidative stress in plant exposed to elevated O₃. Under the background of globe climate change, especially the rapidly increased O₃ and CO₂ in urban areas of China, studies on physiological and biochemical response to these two major greenhouse gases will be necessary for urban greening. Further, response of the antioxidant system in *Ginkgo biloba* leaves to elevated

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O₃ was studied previously (He et al. 2006), and to reveal the interactive effect of the two major greenhouse gases on the antioxidant systems in *Ginkgo biloba* leaves this work was conducted.

Materials and Methods

The experiment was performed at Shenyang Arboretum of Chinese Academy of Sciences (41°46'N, 123°26'E), located in urban area of Shenyang city. Twenty-four-year-old *Ginkgo biloba* seedlings were transplanted into each chamber in late-March 2006 and fumigated with elevated O₃ (80 ppb) and CO₂ (700 ppm), alone or in combination since 15 June for the whole growing season. Additional O₃ was supplied from 8:00 am to 5:00 pm in sunny days. Additional CO₂ was supplied 24 h each day. Chambers with ambient air (O₃, 40 ppb; CO₂, 390 ppm) were set as control and the gases supply and monitor systems were controlled by computer software, as He et al. (2007) described.

Foliar samples, fully expanded leaves without obvious visual damage symptom, were collected at 9:00 am and plunged into liquid N₂ for later biochemical analysis. To calculate dry weight, parallel samples were dried over night at 80°C.

The concentration of H₂O₂ in leaf tissue was determined according to the method of Mukherjee and Choudhuri (1983) with minor modification. Briefly, 0.5 g of leaf segments was ground into fine powder under liquid nitrogen and then homogenized with pre-cooled acetone (5 mL), and finally centrifuged at 10,000 g and 4°C for 10 min. One half milliliter of the supernatant was mixed with 0.5 mL cooled acetone, 0.1 mL 5% Ti (SO₄)₂ and 0.2 mL 19% ammonia. After the precipitate was formed, 1.3 mL reaction mixture was centrifuged at 10,000 g and 4°C for 10 min. The resulting precipitate was dissolved in 5 mL 2 mol L⁻¹ H₂SO₄ and the absorbance of Ti⁴⁺-H₂O₂ compound was recorded at 415 nm. The H₂O₂ concentration was calculated according to a standard curve of H₂O₂.

Malondialdehyde (MDA) and Ascorbate (ASA) were extracted together with 5 mL of 5% (w/v) trichloroacetic acid at about 0°C. The thiobarbituric acid test was used to assay the MDA content in leaves according to the method of Hodges et al. (1999). The concentration of thiobarbituric acid reactive substances (TBARS) was calculated as MDA equivalents using the extinction coefficient of 155 mmol mol⁻¹ cm⁻¹ for malondialdehyde and expressed on a dry weight (DW) basis. Ascorbate analysis was performed on a Waters Alliance 2695 HPLC system and detected with a Waters 996 photodiode array detector according to the method of Liu et al. (1994). The column was eluted at 1 mL min⁻¹ with 0.5% (w/v) NH₄H₂PO₄

solution at pH 2.5. AsA was quantified at its absorption maximum of 254 nm.

Soluble protein and enzymes superoxide dismutase (SOD) and catalase (CAT) was extracted with potassium phosphate buffer (PH 7.8) containing 1 mmol mol⁻¹ EDTA and 2% polyvinylpyrrolidone (PVP) on ice. The supernatant was transferred to a new tube and kept at 4°C for immediate analysis. SOD activity was assayed by the inhibition of photochemical reduction of nitro blue tetrazolium (NBT) according to the method of Krivosheeva et al. (1996). CAT activity was determined by the decrease of absorbance of H₂O₂ at 240 nm in 1 min (Aebi and Lester 1984). Soluble protein content was assayed following the method of Song et al. (2004) using bovine serum albumin (BSA) as a standard.

Ascorbate peroxidase (APX), dehydroascorbate reductase (DHAR) and glutathione reductase (GR) were extracted together and immediately analyzed as described by Jin et al. (2003) with minor modification. APX activity was determined by monitoring the decrease in A280 (absorption coefficient 2.8 mmol mol⁻¹ cm⁻¹) for 1 min in 1 mL of a reaction mixture containing 0.8 mL of 0.5 mmol mol⁻¹ ASA in phosphate buffer (pH 7.0), 0.1 mL of 2 mmol mol⁻¹ H₂O₂ and 0.1 mL of crude enzyme. For the detection of DHAR activity, the reaction mixture contained phosphate buffer (pH 7.0) 0.7 mL, reduced glutathione (GSH) 50 mmol mol⁻¹ 0.1 mL in the phosphate buffer (pH 7.0), 2 mmol mol⁻¹ DHA 0.1 mL, and crude enzyme 0.1 mL. The formation of ascorbate was recorded as the increase in A280 for 1 min after the reaction was started. GR activity was determined by following the consumption of NADPH at 340 nm (absorption coefficient 6.2 mmol mol⁻¹ cm⁻¹) for 1 min in 1 mL of a reaction mixture containing 0.82 mL of 1 mmol mol⁻¹ oxidized glutathione (GSSG), 0.08 mL of 4 mmol mol⁻¹ NADPH in phosphate buffer (pH 7.6), and 0.1 mL of enzyme extract.

Ascorbic acid (ASA, sodium salt), dehydroascorbic acid (DHA), glutathione (GSH), glutathione in oxidized form (GSSG), NADPH, PVP and Triton-X100 were all bought from Sigma and the absorption records were all detected with a Shimadzu UV-1601 spectrophotometer. The results presented are the means (n = 3–6) of all the measurements. Comparisons between means were conducted using the SPSS 10.0 computer package (SPSS Inc. 1999) for all sets of data, by one way ANOVA and the *p* = 0.05 level of error.

Results and Discussion

Both of H₂O₂ and MDA contents were increased significantly by high-ozone exposure for 40 and 80 days (Fig. 1).

Fig. 1 H_2O_2 content (a) and MDA content (b) in *Ginkgo biloba* leaves after 40 and 80 days exposure to elevated O_3 (EO), elevated CO_2 (EC), elevated $\text{O}_3 + \text{CO}_2$ (EOC), and ambient air (AA). Values are means \pm standard error of 3–6 independent determination

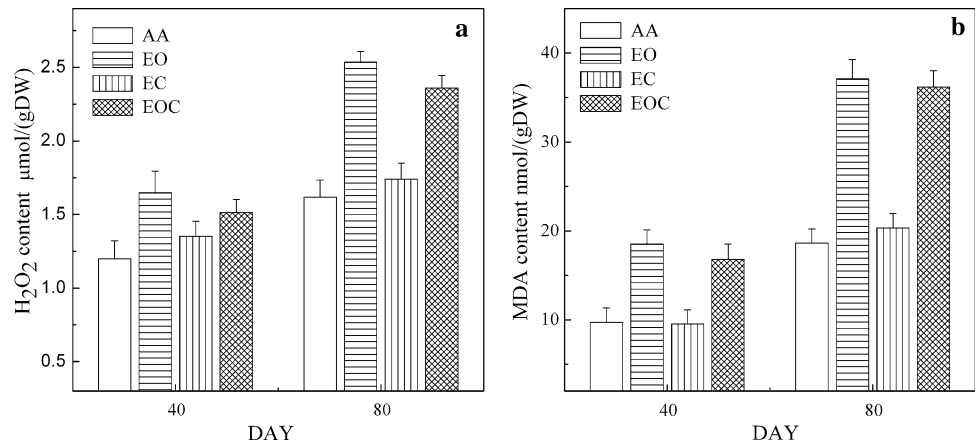


Fig. 2 SOD and CAT activity (a, b) in *Ginkgo biloba* leaves after 40 and 80 days exposure to elevated CO_2 (EC), elevated O_3 (EO), elevated $\text{O}_3 + \text{CO}_2$ (EOC), and ambient air (AA). Values are means \pm standard error of 3–6 independent determination

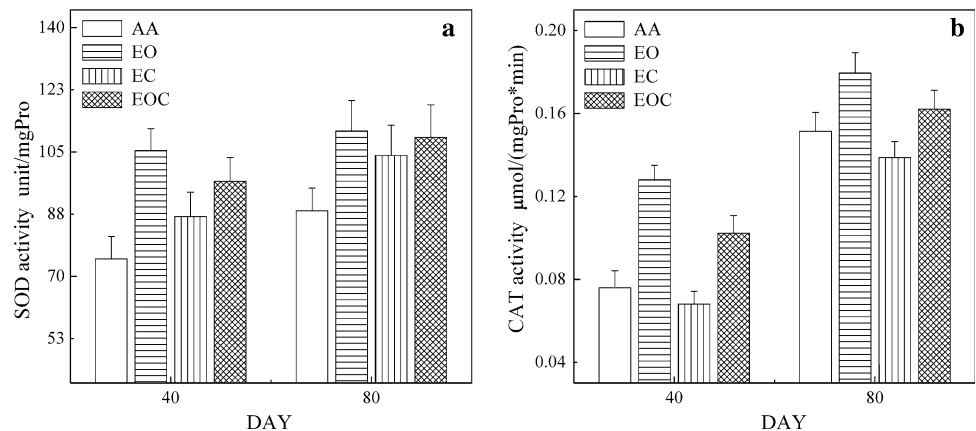
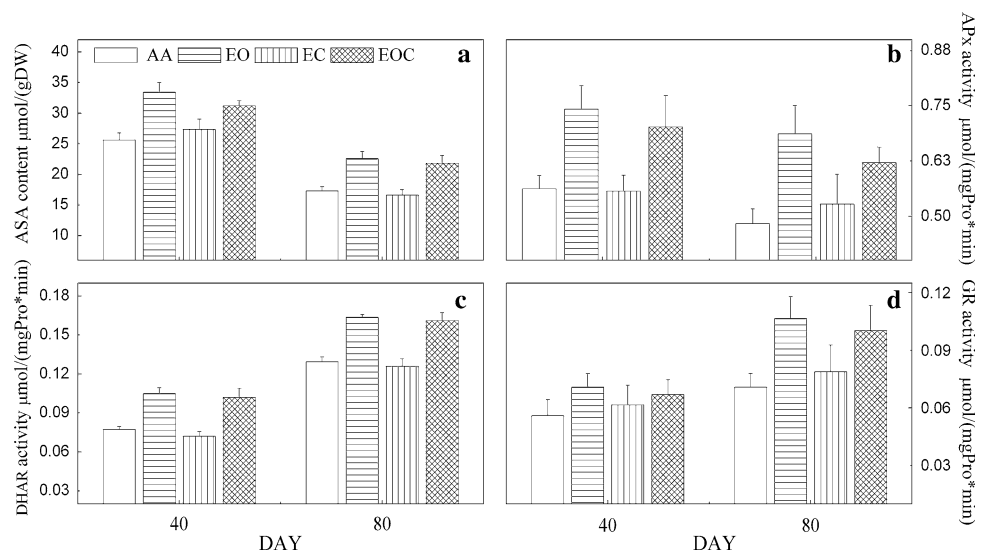


Fig. 3 ASA content (a) and the enzyme activities of APX, DHAR and GR in *Ginkgo biloba* leaves after 40 and 80 days exposure to elevated O_3 (EO), elevated CO_2 (EC), elevated $\text{O}_3 + \text{CO}_2$ (EOC) and ambient air (AA). Values are means \pm standard error of 3–6 independent determination



The specific activities of enzymes in antioxidant systems (Figs. 2, 3b–d) were also significantly enhanced except for GR on the 40th day (Fig. 3d), and so was the content of ascorbate (Fig. 3a) in *Ginkgo biloba* leaves. It suggested that the increase in activity of antioxidant enzymes was an acclimation effect, but the increase was not sufficient to

balance the ROS production, which resulted in the increase in lipid peroxidation as shown by MDA content.

Whether elevated CO_2 would enhance the defense systems in plants is still contradictory. In our research, elevated CO_2 did not significantly change the H_2O_2 and MDA content (Fig. 1). Moreover, the defense system

including the major ROS scavenging enzymes, CAT and APX (Figs. 2b, 3b), and the important small molecular antioxidant ASA (Fig. 3a) did not change significantly either. It may implicate that seedlings exposed to elevated CO₂ produce less ROS and did not need a higher-level antioxidant system. As reported by Karnosky (2003) that, in long term, this could lead to a depressed antioxidant status in plants which would decrease the antioxidant capacity when plants met other stresses.

The effect of simultaneous exposure to both high CO₂ and high O₃ were very similar to exposure to elevated O₃ alone. H₂O₂ and MDA contents of *Ginkgo biloba* leaves (Fig. 1) were still significantly increased, but to a little less extent. Moreover, the specific activity of assayed antioxidant enzymes and ASA content in leaves (Figs. 2, 3) were lower than that exposed to high O₃ alone. It seemed that high CO₂ did not change the response of *Ginkgo biloba* to high O₃ alone. However, increased H₂O₂ and MDA content by high O₃ were mitigated to some extent by high CO₂. Although the mitigating effect may not be statistically significant, the trend was very clearly shown in most measured indexes. These results suggested that high CO₂ perhaps reduced O₃-induced oxidative stress through reducing ROS production (Fig. 1a) which was mainly attributed to decreased O₃ flux into leaf apoplast. As a result, the damage to the cell membrane was alleviated (Fig. 1b) in comparison to exposure of elevated O₃ alone. The lower antioxidant status (Figs. 2, 3) under combined treatment confirmed the lower demand of ROS scavenging in comparison to elevated O₃ alone.

Differences in the response to exposures were discernible between the 40th and 80th day. Higher level of H₂O₂ and MDA content (Fig. 1) on the 80th day perhaps indicated the seedlings' natural senescence and/or the accumulative effect of O₃ exposure with or without elevated CO₂. This view was supported by the higher ASA content (Fig. 3a) on the 40th day. On the 80th day the specific activity of enzymes assayed in Ascorbate-Glutathione cycle was more stimulated by high O₃ with or without high CO₂, especially GR (Fig. 3d), of which was significantly stimulated by 50.6% or 41.8%, respectively. In correspondence, on the 40th day the enhancement of SOD and CAT activity (Fig. 2) by O₃ exposure with or without elevated CO₂ was much stronger than that on the 80th day, 40.7% and 68.9%, 29.1% and 34.7%, respectively in comparison with that under ambient air. It revealed that different compartments of the antioxidant system in *Ginkgo biloba* leaves responded to elevated O₃ or/and CO₂ distinctively at different periods in the growing season. To reveal the defense mechanism of plants against oxidative stresses at cellular level further researches will be needed.

In conclusion, the enhancement of antioxidant system in *Ginkgo biloba* seedlings exposed to elevated O₃ alone was

not sufficient to balance the increased H₂O₂ and MDA production. In the air with elevated CO₂ alone, it is not necessary to enhance the defense system against oxidative stress, however, in the case plant exposed to more oxidative environment like elevated O₃, the elevated CO₂ showed an effect in mitigating the oxidative stress.

Acknowledgments This work was funded by the National Natural Science Foundation of China Important Project 90411019, the Foundation of Knowledge Innovation Program of Chinese Academy of Sciences kzcx3-sw-43.

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